

Biomimetic Total Synthesis of (\pm) -Doitunggarcinone A and (+)-Garcibracteatone

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Supporting Information

ABSTRACT: A full account of our oxidative radical cyclization approach to the synthesis of garcibracteatone and doitunggarcinone A is presented. This includes the first enantioselective synthesis of garcibracteatone, which allowed the absolute configuration of the natural compound to be determined. The first synthesis of doitunggarcinone A is also described, which confirms our reassignment of the relative configuration of this molecule. Novel syntheses of mono-

terpene fragments used to construct the target molecules are also reported.

■ INTRODUCTION

Polycyclic polyprenylated acylphloroglucinols (PPAPs) are a large and structurally diverse family of plant-derived natural products. The chemistry and biology of these compounds has been summarized in several recent reviews, and there are many successful total syntheses of PPAP natural products.² Most PPAPs contain a bicyclo[3.3.1]nonane ring system, which is proposed to be biosynthesized via electrophilic cyclizations of monocyclic polyprenylated acylphloroglucinols.3 However, we believe that many of the more complex PPAP natural compounds, such as garcibracteatone (1),⁴ nemorosonol (3),⁵ doitunggarcinone A (2), doitunggarcinone B (4), ialibinone A (5), ialibinone B (6), and peroxysampsone A (7)⁸ are formed in nature via oxidative radical cyclizations (Figure 1). Porco has previously shown that oxidative radical cyclizations can be used to build up complex unnatural PPAP structures, and we have previously used an oxidative radical cyclization approach in the synthesis of (\pm) -ialibinones A (5) and B (6). Herein, we give a full account of our biomimetic syntheses of (+)-garcibracteatone $(1)^{11}$ and (\pm) -doitunggarcinone A (2).

Garcibracteatone and nemorosonol were co-isolated from Garcinia bracteata, while doitunggarcinones A and B were coisolated from Garcinia propingua, thus suggesting a close biogenetic relationship within these pairs of natural products. Our proposed biosynthesis of garcibracteatone, nemorosonol, and doitunggarcinones A and B is outlined in Scheme 1. The starting point for the radical cyclization pathways are the dearomatized compounds 8 and 9, which could be formed by multiple prenylations of acylphloroglucinol derivatives. Compound 8 is a possible diastereomer of weddellianone A, 12 a known acylphloroglucinol natural product isolated from Clusia weddelliana, but whose relative and absolute configurations have not been fully established. Single electron oxidation of compound 8 or 9 would give rise to a stabilized α -diketo radical, which could undergo successive 7-endo-trig and 5-exo-

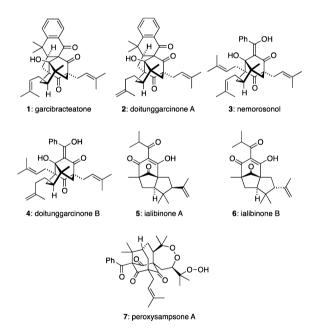


Figure 1. PPAP natural products possibly biosynthesized by radical cyclizations.

trig cyclizations followed by hydrogen atom abstraction to give nemorosonol (3) or doitunggarcinone B (4). Alternatively, a second 5-exo-trig cyclization followed by an aromatic radical substitution reaction would give rise to garcibracteatone (1) or doitunggarcinone A (2). Although this biosynthetic pathway constitutes a highly complex sequence of reactions, we reasoned that the compact nature of the radical intermediates

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Scheme 1. Proposed Biosynthesis of Garcibracteatone, Nemorosonol, and Doitunggarcinones A and B

might render the cyclizations predisposed to occur under nonenzymatic conditions in a biomimetic synthesis.

■ RESULTS AND DISCUSSION

We initially investigated the feasibility of a radical cyclization approach to PPAP synthesis through some model studies using the dearomatized phloroglucinol 13 as a simplified analogue of 8 or 9 (Scheme 2). The use of a 2-methyl-1-butene substituent at C-5 instead of the natural lavandulyl side chain simplified the system considerably, and the presence of allyl groups at C-3 and C-5 instead of prenyl groups would be expected to reduce the possibility of competing 5-exo-trig cyclizations (as observed in our biomimetic synthesis of ialibinones A and B). Thus, 2,4,6-trihydroxybenzophenone $(10)^{13}$ was allylated at C-3 and C-5 using allyl bromide in aqueous KOH to give 11 in 22% yield. Dearomatization of 11 with NaH and 4-iodo-2-methyl-1butene (12)¹⁴ in DMF gave 13 in good yield. Oxidation of 13 using Mn(OAc)₃¹⁵ in EtOH then gave **16** as the only isolable product, in low yield of 5%. Presumably 16 was formed via oxidative 7-endo-trig and 5-exo-trig cyclizations common to the proposed biosynthesis outlined in Scheme 1 to form the α diketo radical 14. However, this radical apparently underwent an unexpected final 6-endo-trig cyclization onto the C-5 allyl group to give the secondary radical 15, followed by a terminating hydrogen atom abstraction step to give 16. We had hoped that the α -diketo radical intermediate 14 might undergo hydrogen atom abstraction itself, thereby forming the

Scheme 2. Model Oxidative Cyclization Studies

polycyclic ring system common to nemorosonol (3) and doitunggarcinone B (4).

Although the oxidative radical cyclization of 13 did not proceed as planned, we were encouraged that the initial 7-endotrig and 5-exo-trig cyclizations had apparently worked to some degree. We reasoned that replacement of the allyl groups of 13 with prenyl groups might encourage a diketo radical such as 14 to undergo a second 5-exo-trig cyclization, rather than a 6-endotrig cyclization. A terminating intramolecular aromatic radical substitution reaction might then form the polycylic framework of garcibracteatone (1) and doitunggarcinone A (2). Thus, 2,4,6-trihydroxybenzophenone (10) was reacted with prenyl bromide in aqueous KOH to give 17 in 34% yield (Scheme 3). 16 Dearomatization of 17 with NaH and 4-iodo-2-methyl-1butene (12) in DMF then gave 18 in 54% yield. Treatment of 18 with Mn(OAc)₃ and Cu(OAc)₂ in AcOH gave 19 in 25% yield via a radical cyclization cascade that formed the complete garcibracteatone/doitunggarcinone A framework in one step, presumably via the mechanism outlined in Scheme 1. The low yield of 25% is compensated by the formation of four carboncarbon bonds and five stereocenters in the reaction, with complete control of relative stereochemistry. The structure of 19 was elucidated by NMR studies, with the ¹H and ¹³C NMR spectra showing close similarity to those of natural garcibracteatone (1) and doitunggarcinone A (2). The structure of 19 was later confirmed by X-ray crystallographic studies.¹⁷ No products containing the nemorosonol/doitunggarcinone B ring system were observed during either of these model studies. Screening of a variety of alternative one-electron oxidants, such as PhI(OAc)2, PhI(OTf)2, and CAN, failed to generate any detectable quantities of 19.

In order to apply the radical cyclization cascade to a full total synthesis of (\pm) -garcibracteatone $((\pm)$ -1), we now needed to alkylate 17 with (\pm) -lavandulyl iodide $((\pm)$ -23), which could be obtained by iodination of (\pm) -lavandulol $((\pm)$ -22, Scheme 4). Although (\pm) -lavandulol is commercially available, we had

Scheme 3. Model Oxidative Cyclization Studies

Scheme 4. Synthesis of (\pm) -Lavandulyl Iodide

difficulty obtaining it cheaply, and we therefore developed an efficient and economical synthesis from ethyl 3,3-dimethylacrylate (20). Deprotonation of 20 with LDA in THF followed by α -alkylation with prenyl bromide gave (\pm)-21 in high yield, with concomitant shift of the alkene from the α , β to the β , γ position. The ethyl ester of (\pm)-21 was then reduced using LiAlH₄ to give (\pm)-lavandulol ((\pm)-22) in excellent yield. ¹⁸ Iodination of (\pm)-22 under standard conditions then gave (\pm)-23 in 86% yield.

Alkylation of 17 with (\pm) -23 in DMF with NaH as the base gave (\pm) -24 in 29% yield as a complex mixture of enol tautomers and diastereomers, as the relative C-1/C-5 stereochemistry (using garcibracteatone numbering) was not controlled during the reaction (Scheme 5). The yield for the alkylative dearomatization was low due to the sterically hindered nature of the alkyl iodide and competing E2 elimination. However, oxidative cyclization of (\pm) -24 using Mn(OAc)₃ and Cu(OAc)₂ in AcOH gave (\pm) -garcibracteatone $((\pm)$ -1) in 14% yield and (\pm) -5-epi-garcibracteatone $((\pm)$ -25) in 8% yield, which were separable by flash chromatography on silica gel. The mechanism of the oxidative cyclization of (\pm) -24 presumably follows the radical cascade outlined in Scheme 1. X-

Scheme 5. Synthesis of (\pm) -Garcibracteatone

ray crystallographic studies of (\pm) -1 and (\pm) -25 confirmed their structures, and comparison with the NMR data for the natural material allowed us to confirm the previously undefined C-5 relative configuration of (\pm) -garcibracteatone to be as shown in Scheme 5.

Our next target was an enantioselective synthesis of garcibracteatone, which would require access to enantiopure lavandulyl iodide. We therefore synthesized (-)-23 according to Scheme 6 using a chiral auxiliary approach. Alkylation of (-)-26 with prenyl bromide and NaHMDS as the base gave (+)-27 in 66% yield and 15:1 dr. Reduction of (+)-27 with LiBH₄ gave (-)-22, which was iodinated to give (-)-23 under

Scheme 6. Enantioselective Synthesis of (-)-Lavandulyl Iodide

standard conditions. This represents a practical synthesis of enantiopure lavandulol and lavandulyl iodide that is significantly shorter than existing methods.²¹

With (-)-lavandulyl iodide in hand, we were able to synthesize the dearomatized phloroglucinol (-)-24 in enantio-enriched form and then (+)-garcibracteatone ((+)-1) and (-)-5-epi-garcibracteatone ((-)-25) via oxidative radical cyclization (Scheme 7). Naturally occurring (-)-garcibractea-

Scheme 7. Enantioselective Synthesis of (+)-Garcibracteatone

tone has a very low specific rotation ($[\alpha]^{25}_D$ –1 (c 1.0, CHCl₃)), and our synthetic (+)-garcibracteatone had a similarly low value ($[\alpha]^{25}_D$ +2 (c 1.0, CHCl₃)). Given these low magnitudes, the signs of the specific rotations of natural and synthetic garcibracteatone are probably within the bounds of experimental error and do not, by themselves, allow the determination of the absolute configuration of natural garcibracteatone. However, we were fortunate to have access to a sample of natural garcibracteatone, and comparison of chiral HPLC traces between this and our synthetic material conclusively showed that we had made the enantiomer of the natural product. We can therefore assign the absolute stereochemistry of natural (–)-garcibracteatone to be as shown in Scheme 7.

Shortly after our synthesis of garcibracteatone (1) had been completed, the isolation of doitunggarcinone A (2) and doitunggarcinone B (4) from *Garcinia propinqua* was reported. These natural products have very similar structures to garcibracteatone (1) and nemorosonol (3), differing only in the position of the carbon—carbon double bond in the C-5 alkyl chain. In addition, they were originally assigned as having the opposite relative configuration at C-5 compared to garcibracteatone (1) and nemorosonol (3). However, close inspection of the ¹H and ¹³C NMR data for doitunggarcinone A (2) showed very good correlation with the NMR spectra of our synthetic garcibracteatone (1), whereas comparison with the NMR spectra of synthetic 5-epi-garcibracteatone (25) showed significant differences. Similarly, the NMR data for doitung-

garcinone B (4) are almost identical to those of nemorosonol (3). We therefore suggest that structures 2 and 4 represent the correct stereochemical assignment for doitunggarcinones A and B, respectively, as shown in Figure 2.

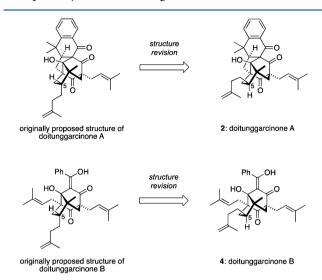


Figure 2. Suggested structure revision of doitunggarcinones A and B.

We sought to confirm our structural reassignment of doitunggarcinone A (2) through a total synthesis of the racemic compound. For this we first needed to synthesize alkyl iodide (\pm) -30, which was achieved according to Scheme 8.

Scheme 8. Synthesis of Alkyl Iodide (±)-30

Alkylation of ethyl 3,3-dimethylacrylate (20) with 4-iodo-2-methyl-1-butene (12) and LDA gave (\pm)-28, which was reduced to give alcohol (\pm)-29. Iodination with I₂ and PPh₃ then gave (\pm)-30 in good yield over the three steps. Unfortunately, an enantioselective synthesis of 30 following a similar strategy to that outlined in Scheme 6 proved elusive, as alkylation of (–)-26 with 4-iodo-2-methyl-1-butene (12) was not possible.

Alkylative dearomatization of 17 with (\pm) -30 formed 31 in 33% yield, and then oxidative cyclization gave (\pm) -doitunggarcinone A $((\pm)$ -2)) in 13% yield and (\pm) -5-epi-doitunggarcinone A $((\pm)$ -32)) in 12% yield (Scheme 9). NMR data for (\pm) -2 fully matched the published data for the natural product. The relative configuration of (\pm) -2 at C-5 was established via observation of a strong NOE between H-5 and H-10, thus

Scheme 9. Synthesis of (\pm) -Doitunggarcinone A

confirming our suggested structure revision. An NOE interaction between H-5 and Me-26 further confirmed the structure of (\pm) -32.

Interestingly, no formation of nemorosonol or doitunggarcinone B was ever observed in our total syntheses of garcibracteatone and doitunggarcinone A. Therefore, attempts were made to carry out a shortened radical cascade reaction with just two cyclization events that would allow synthesis of the nemorosonol/doitunggarcinone B ring system. Thus, deactivation of the prenyl side chains of 17 by hydrogenation gave 33 in 78% yield, which was then alkylated to give the dearomatized phloroglucinol 34 in 37% yield (Scheme 10). However, oxidation of 34 with a variety of oxidants gave complex mixtures of products, with no observed formation of the desired product 35.

Scheme 10. Attempted Synthesis of the Nemorosonol/ Doitunggarcinone B Ring System

CONCLUSION

We have shown that oxidative radical cyclizations are highly effective in the construction of complex PPAP natural products such as garcibracteatone (1) and doitunggarcinone A (2) and previously ialibinones A (5) and B (6). The success of these radical reactions in rapidly building up molecular complexity, with the formation of multiple stereocenters and carbon—carbon bonds in a single operation, perhaps indicates that similar pathways are involved in the biosynthesis of these PPAPs. Furthermore, we believe that there are several more PPAP natural products that could arise via complexity-generating radical cascade cyclizations. For example, the ornate structure of peroxysampsone A (7)⁸ could be formed by oxidative radical cyclization of the related natural product 7-epinemorosone (36) according to Scheme 11. Initial single

Scheme 11. Proposed Biosynthesis of Peroxysampsone A

electron oxidation of 36 could give a stabilized α -diketo radical, which could undergo a 7-exo-trig radical cyclization onto the adjacent C-7 prenyl group to give a tertiary alkyl radical. Trapping of this radical with triplet oxygen, followed by 7-exo-trig cyclization of the resultant peroxyradical species, would then generate the complex ring system of peroxysampsone A (7).

■ EXPERIMENTAL SECTION

General Methods. All chemicals used were purchased from commercial suppliers and used as received. All reactions were performed under an inert atmosphere of N_2 . All organic extracts were dried over anhydrous magnesium sulfate. Thin layer chromatography was performed using aluminum sheets coated with silica gel. Visualization was aided by viewing under a UV lamp and staining with ceric ammonium molybdate stain followed by heating. All R_f values were measured to the nearest 0.01. Flash chromatography was performed using 40–63 μ m grade silica gel. Melting points were recorded on a digital melting point apparatus and are uncorrected. Infrared spectra were recorded using an FT-IR spectrometer as the

neat compounds. High field NMR was recorded using a 600 MHz spectrometer ($^1\mathrm{H}$ at 600 MHz, $^{13}\mathrm{C}$ at 150 MHz). Solvents used for spectra were chloroform unless otherwise specified. $^1\mathrm{H}$ chemical shifts are reported in ppm on the δ -scale relative to TMS (δ 0.0), and $^{13}\mathrm{C}$ NMR are reported in ppm relative to chloroform (δ 77.0). Multiplicities are reported as (br) broad, (s) singlet, (d) doublet, (t) triplet, (q) quartet, and (m) multiplet. All J values were rounded to the nearest 0.1 Hz. ESI high resolution mass spectra were recorded on a Q-TOF mass spectrometer.

Phenyl(2,4,6-trihydroxyphenyl)methanone (10). To a suspension of anhydrous phloroglucinol (11.5 g, 91.2 mmol) in PhNO₂ (90 mL) was added AlCl₃ (48.6 g, 365 mmol) in three portions at rt. The reaction mixture was stirred at rt for 30 min. Benzoyl chloride (11.7 mL, 100 mmol) was added and was heated at 65 °C for 2 h. The reaction mixture was then cooled to rt before being quenched by pouring onto ice—water and extracted with EtOAc (3 × 100 mL). The product was then extracted into 2 M NaOH solution (2×150 mL). The aqueous extracts were neutralized with conc. HCl, and the product was extracted back into EtOAc (3 × 100 mL). The combined organics were washed sequentially with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, $4:1 \rightarrow 2:1$ gradient elution) to yield phenyl(2,4,6-trihydroxyphenyl)methanone 10 (9.80 g, 47%) as a yellow solid. Data for 10: R_f 0.10 (petrol/EtOAc, 2:1); mp 164-167 °C; IR (neat) 3363, 1638, 1594, 1286, 1151, 1056, 818, 697 cm⁻¹; 1 H NMR (600 MHz, acetone- d_6) δ 10.15 (br s, 3H), 7.57 (d, J = 7.2 Hz, 2H), 7.44 (t, J = 7.2 Hz, 1H), 7.36 (t, J = 7.2 Hz, 2H), 5.95 (s, 2H); ¹³C NMR (150 MHz, acetone d_6) δ 199.5, 165.2, 163.6, 142.3, 131.3, 128.6, 128.0, 104.9, 95.6.

(3,5-Diallyl-2,4,6-trihydroxyphenyl)(phenyl)methanone (11). To a solution of 10 (2.29 g, 9.95 mmol) in H₂O (20 mL), was added KOH (1.12 g, 19.9 mmol) at 0 °C. Allyl bromide (1.71 mL, 19.9 mmol) was then added dropwise over 20 min. The reaction mixture was warmed to rt and stirred for a further 1 h. The mixture was acidified with 1 M HCl solution (10 mL) and then extracted with EtOAc (3 \times 20 mL). The combined organics were washed with brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 8:1 as elutant) to give (3,5-diallyl-2,4,6trihydroxyphenyl)(phenyl)methanone 11 (691 mg, 22%) as a yellow solid. Data for 11: R_f 0.29 (petrol/EtOAc, 4:1); mp 83–85 °C; IR (neat) 3468, 3234, 2926, 1620, 1590, 1560, 1205, 1106, 911, 697 cm⁻¹ H NMR (600 MHz, CDCl₃) δ 8.92 (s, 1H), 7.64 (dd, J = 8.2, 1.3 Hz, 2H), 7.59 (t, J = 7.5 Hz, 1H), 7.52 (t, J = 7.5 Hz, 2H), 5.98 (s, 1H), 6.00-5.92 (m, 2H), 5.15 (dq, J = 26.7, 1.6 Hz, 2H), 5.13 (dq, J = 19.6, 1.6 Hz, 2H), 3.41 (t, J = 1.6 Hz, 2H), 3.40 (t, J = 1.6 Hz, 2H). 13 C NMR (150 MHz, CDCl₃) δ 197.9, 161.0, 158.0, 139.9, 136.1, 132.2, 129.2, 127.8, 116.0, 104.6, 27.0; HRMS-ESI (m/z) calculated for $C_{19}H_{19}O_4 [M + H]^+ 311.1278$, found 311.1278.

4,6-Diallyl-2-benzoyl-3,5-dihydroxy-6-(3-methylbut-3-en-1yl)cyclohexa-2,4-dienone (13). To a solution of 11 (400 mg, 1.29 mmol) in anhydrous DMF (8 mL) was added NaH (60% dispersion in mineral oil, 155 mg, 3.87 mmol) at rt. The mixture was stirred at rt for 5 min before 4-iodo-2-methyl-1-butene 12 (502 mg, 2.56 mmol) was added at rt. The reaction mixture was stirred at rt for 1 h. The mixture was quenched with 1 M HCl solution (10 mL) and extracted with EtOAc (3×15 mL). The combined organics were washed sequentially with H_2O (2 × 30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, $6:1 \rightarrow 4:1$ gradient elution) to give 4,6-diallyl-2-benzoyl-3,5-dihydroxy-6-(3-methylbut-3en-1-yl)cyclohexa-2,4-dienone 13 (371 mg, 76%) as a yellow solid. Data for 13: R_f 0.14 (petrol/EtOAc, 4:1); mp 97-101 °C; IR (neat) 3167, 2878, 1648, 1198, 890, 696 cm⁻¹; NMR spectra showed a complex mixture of tautomers; HRMS-ESI (m/z) calculated for $C_{24}H_{27}O_4 [M + H]^+$ 379.1904, found 379.1907.

7-Allyl-5a-benzoyl-2a1-hydroxy-8a-methyloctahydro-1H-2a,7-methanoacenaphthylene-6,9(2H)-dione (16). To a solution of Mn(OAc) $_3$ (H $_2$ O) $_2$ (212 mg, 0.79 mmol) in degassed EtOH (1 mL) was added 13 (150 mg, 0.396 mmol) in degassed EtOH (6 mL) at rt.

The reaction mixture was stirred at rt for 3 h. The mixture was quenched with H_2O (15 mL) and was extracted with EtOAc (3 × 20 mL). The combined organics were washed sequentially with H₂O (30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 10:1 as elutant) to give 7-allyl-5abenzoyl-2a1-hydroxy-8a-methyloctahydro-1H-2a,7-methanoacenaphthylene-6,9(2H)-dione 16 (7 mg, 5%) as a white crystalline solid. Data for **16**: R_f 0.35 (petrol/EtOAc, 4:1); mp 115–117 °C; IR (neat) 3528, 2932, 1736, 1707, 1650, 1243, 1072, 914, 689 cm⁻¹; ¹H NMR (600 MHz, CDCl₂) δ 7.56 (dd, I = 8.2, 1.3 Hz, 2H), 7.52 (t, I = 7.5 Hz, 1H), 7.40 (t, J = 7.8 Hz, 2H), 5.71 (m, 1H), 5.09 (d, J = 4.1 Hz, 1H), 5.06 (s, 1H), 4.71 (s, 1H), 2.57 (dd, *J* = 14.4, 7.0 Hz, 1H), 2.39–2.30 (m, 2H), 2.28 (ddd, J = 12.7, 4.3, 2.2 Hz, 1H), 1.97 (m, 3H), 1.86 (td, J = 12.1, 6.7 Hz, 1H), 1.74 (ddd, <math>J = 12.5, 9.5, 3.1 Hz, 1H), 1.58 (t, J = 12.5, 9.5, 3.1 Hz, 1H)7.1 Hz, 1H), 1.56-1.54 (m, 1H), 1.54-1.51 (m, 1H), 1.43 (ddd, J =13.5, 9.5, 6.7 Hz, 1H), 1.18 (s, 3H), 0.95-0.82 (m, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 210.8, 207.2, 204.6, 137.9, 133.4, 132.1, 128.3, 127.9, 118.7, 80.6, 66.5, 65.7, 62.8, 43.3, 40.5, 39.0, 35.5, 31.3, 30.6, 28.0, 23.3, 19.8; HRMS-ESI (m/z) calculated for $C_{24}H_{27}O_4$ $[M + H]^+$ 379.1904, found 379.1901.

Phenyl(2,4,6-trihydroxy-3,5-bis(3-methylbut-2-en-1-yl)phenyl)methanone (17). To a solution of 10 (10.0 g, 43.4 mmol) in H₂O (80 mL) was added KOH (4.89 g, 86.9 mmol) at 0 °C. Prenyl bromide (10.0 mL, 86.9 mmol) was then added dropwise over 20 min at 0 °C. The reaction mixture was stirred at 0 °C for a further 1 h. The reaction mixture was acidified with 1 M HCl solution (40 mL) and then extracted with EtOAc (3 × 100 mL). The combined organics were washed with brine (200 mL), dried over MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, $20:1 \rightarrow 10:1$ gradient elution) to give phenyl(2,4,6-trihydroxy-3,5-bis(3-methylbut-2-en-1-yl)phenyl)methanone 17 (5.35 g, 34%) as a yellow solid. Data for 17: R_f 0.45 (petrol/EtOAc, 4:1); mp 76-82 °C; IR (neat) 3360, 2912, 1618, 1560, 1427, 1325, 1098, 694 cm $^{-1}$; ¹H NMR (600 MHz, CDCl₃) δ 8.91 (s, 2H), 7.65-7.62 (m, 2H), 7.59-7.55 (m, 1H), 7.52-7.48 (m, 2H), 6.35 (s, 1H), 5.22 (t, J = 7.1 Hz, 2H), 3.34 (d, J = 7.0 Hz, 4H), 1.78 (s, 6H), 1.74 (s, 6H); 13 C NMR (150 MHz, CDCl₃) δ 198.0, 161.0, 157.6, 140.3, 135.0, 132.0, 129.0, 127.9, 121.8, 106.3, 104.5, 25.8, 21.8, 17.9.

2-Benzoyl-3,5-dihydroxy-4,6-bis(3-methylbut-2-en-1-yl)-6-(3-methylbut-3-en-1-yl)cyclohexa-2,4-dienone (18). To a solution of 17 (454 mg, 1.24 mmol) in anhydrous DMF (5 mL) was added NaH (60% dispersion in mineral oil, 149 mg, 3.72 mmol) at rt. The mixture was stirred at rt for 5 min. 4-Iodo-2-methyl-1-butene 12 (0.29 mL, 2.48 mmol) was then added at rt. The reaction mixture was stirred at rt for 1 h. The mixture was quenched with 1 M HCl solution (10 mL) and extracted with EtOAc (3 × 15 mL). The combined organics were washed sequentially with H_2O (2 × 30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, $6:1 \rightarrow 4:1$ gradient elution) to give 2-benzoyl-3,5dihydroxy-4,6-bis(3-methylbut-2-en-1-yl)-6-(3-methylbut-3-en-1-yl)cyclohexa-2,4-dienone 18 (293 mg, 54%) as a viscous yellow oil. Data for 18: R_f 0.23 (petrol/EtOAc, 4:1); IR (neat) 2914, 1647, 1446, 1370, 1186, 693 cm⁻¹; NMR spectra showed a complex mixture of tautomers; HRMS-ESI (m/z) calculated for $C_{28}H_{35}O_4$ $[M + H]^+$ 435.2530, found 435.2528.

2a1-Hydroxy-4,4,12a-trimethyl-11-(3-methylbut-2-en-1-yl)-3,3a,4,11,12,12a-hexahydro-1H-2a,11-methanocyclopenta-[3,A]indeno[1,7a-b]naphthalene-9,10,13(2H,2a1H)-trione (19). To a solution of Mn(OAc) $_3$ (H $_2$ O) $_2$ (252 mg, 0.94 mmol) and Cu(OAc) $_2$ (H $_2$ O) (86 mg, 0.45 mmol) in degassed AcOH (2 mL) was added 18 (195 mg, 0.45 mmol) in degassed AcOH (8 mL) at rt. The reaction mixture was stirred at rt for 3 h. The mixture was quenched with H $_2$ O (15 mL) and extracted with EtOAc (3 × 20 mL). The combined organics were washed sequentially with H $_2$ O (30 mL), sat. NaHCO $_3$ solution (30 mL) and brine (30 mL), dried over anhydrous MgSO $_4$, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on SiO $_2$ (petrol/EtOAc, 10:1 as elutant) to

give 2a1-hydroxy-4,4,12a-trimethyl-11-(3-methylbut-2-en-1-yl)-3,3a,4,11,12,12a-hexahydro-1*H*-2a,11 methanocyclopenta[3,4]indeno-[1,7a-b]naphthalene-9,10,13(2H,2a1H)-trione 19 (50 mg, 25%) as a white crystalline solid. Data for 19: R_f 0.42 (petrol/EtOAc, 4:1); mp 204-206 °C; IR (neat) 3468, 2949, 1736, 1707, 1663, 1599, 1118, 765 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.70 (dd, J = 7.6, 1.3 Hz 1H), 7.54 (td, J = 7.6, 1.3 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.35 (d, J = 7.6Hz, 1H), 5.05 (t, I = 7.2 Hz, 1H), 2.85 (s, 1H), 2.68 (dd, I = 9.8, 7.7 Hz, 1H), 2.37 (t, J = 12.1 Hz, 1H), 2.28 (d, J = 7.3 Hz, 2H), 2.26–2.23 (m, 1H), 2.06 (dd, J = 11.5, 7.7 Hz, 1H), 1.98-1.93 (m, 1H), 1.82-1.931.71 (m, 2H), 1.78 (d, J = 13.9 Hz, 1H), 1.68 (d, J = 13.9 Hz, 1H), 1.65 (s, 3H), 1.57 (s, 3H), 1.49 (s, 3H), 1.36 (s, 3H), 1.11 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 213.4, 203.4, 200.2, 150.3, 136.3, 134.3, 133.7, 126.9, 126.4, 123.5, 118.8, 90.9, 72.8, 68.5, 63.1, 57.0, 46.0, 44.7, 39.5, 37.2, 29.9, 29.0, 26.2, 25.9, 25.1, 24.5, 22.9, 17.9; HRMS-ESI (*m*/ z) calculated for $C_{28}H_{33}O_4$ [M + H]⁺ 433.2373, found 433.2372

 (\pm) -Ethyl 5-methyl-2-(prop-1-en-2-yl)hex-4-enoate $((\pm)$ -21). To a solution of LDA (2.0 M in heptane, 46.8 mL, 93.6 mmol) and anhydrous THF (80 mL) was added ethyl 3,3-dimethyl acrylate 20 (10.0 g, 78 mmol) in anhydrous THF (12 mL) dropwise at −78 °C. The mixture was stirred at -78 °C for 15 min. Prenyl bromide (9.90 mL, 85.8 mmol) was then added at -78 °C. The reaction mixture was stirred for a further 1 h before gradual warming to rt. The mixture was quenched with satd NH₄Cl solution (100 mL) and extracted with Et₂O (3 \times 100 mL). The combined organics were washed with brine (200 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 20:1 as elutant) to give (±)-ethyl 5-methyl-2-(prop-1en-2-yl)hex-4-enoate (\pm) -21 (13.3 g, 87%) as a colorless oil. Data for (\pm)-21: R_f 0.65 (petrol/EtOAc, 4:1); IR (neat) 2976, 1733, 1647, 1179, 1150, 895 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 5.03 (t, J = 7.1Hz, 1H), 4.91-4.86 (m, 2H), 4.13 (qd, J = 7.1, 1.8 Hz, 2H), 3.01 (t, J= 7.7 Hz, 1H), 2.51 (dt, I = 15.1, 7.8 Hz, 1H), 2.26 (dt, I = 14.3, 7.0 Hz, 1H), 1.76 (s, 3H), 1.68 (s, 3H), 1.62 (d, J = 11.5 Hz, 3H), 1.24 (t, J = 7.1 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 173.5, 142.5, 133.4, 121.2, 113.4, 60.4, 53.3, 29.0, 25.7, 20.5, 17.8, 14.2.

 (\pm) -5-Methyl-2-(prop-1-en-2-yl)hex-4-en-1-ol $((\pm)$ -22). To a solution of LiAlH $_4$ (5.60 g, 148 mmol) in Et $_2$ O (200 mL) was added 21 (13.2 g, 67.5 mmol) in Et₂O (60 mL) dropwise over 20 min at 0 °C. The reaction mixture was warmed to rt and stirred for 1 h. The mixture was then cooled to 0 °C, quenched by careful dropwise addition of H₂O (5.6 mL), and stirred at rt for 5 min. Then 15% NaOH solution (5.6 mL) was added, and the mixture was stirred at rt for a further 5 min before H₂O (16.8 mL) was added. The mixture was filtered and extracted thoroughly with Et2O, and the filtrate was concentrated in vacuo to yield (±)-5-methyl-2-(prop-1-en-2-yl)hex-4en-1-ol (\pm)-22 (9.90 g, 95%) as a colorless oil which was used in the next step without further purification. Data for (\pm) -22: R_f 0.39 (petrol/EtOAc, 4:1); IR (neat) 3351, 2916, 1646, 1440, 1376, 1038, 888 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 5.08 (tt, I = 7.2, 1.2 Hz, 1H), 4.94-4.91 (m, 1H), 4.82 (d, J = 0.6 Hz, 1H), 3.57 (dt, J = 11.3, 5.8 Hz, 1H), 3.50 (dt, J = 8.7, 3 Hz, 1H) 2.28 (qd, J = 7.6, 5.2 Hz, 1H), 2.11 (dt, J = 14.6, 7.3 Hz, 1H), 2.04 (dt, J = 14.6, 7.1 Hz, 1H), 1.70 (s, 3H), 1.69 (s, 3H), 1.61 (s, 3H), 1.43 (t, J = 5.3 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 145.4, 132.7, 122.0, 113.1, 63.6, 49.9, 28.4, 25.7,

(±)-3-(lodomethyl)-2,6-dimethylhepta-1,5-diene ((±)-23). To a solution of PPh₃ (18.5 g, 70.6 mmol) and imidazole (4.80 g, 70.6 mmol) in CH₂Cl₂ (200 mL) was added I₂ (17.9 g, 70.6 mmol) at 0 °C. The mixture was stirred at 0 °C for 15 min. Then 22 (9.90 mL, 64.2 mmol) in CH₂Cl₂ (30 mL) was added dropwise at 0 °C. The reaction mixture was stirred at rt for 2 h. The mixture was quenched with Na₂S₂O₃ solution (36 g in 200 mL of H₂O) and stirred at rt for 10 min. The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (200 mL). The combined organics were washed with brine (200 mL), dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on SiO₂ (neat petrol as elutant) to give (±)-3-(iodomethyl)-2,6-dimethylhepta-1,5-diene (±)-23 (14,6 g, 86%) as a pale orange oil. Data for (±)-23: R_f 0.70 (neat petrol); IR (neat) 2968,

2913, 1647, 1439, 1375, 1187, 893 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 5.03 (tt, J = 7.9, 1.3 Hz, 1H), 4.91–4.89 (m, 1H), 4.75 (s, 1H), 3.28 (dd, J = 9.8, 5.8 Hz, 1H), 3.19 (dd, J = 9.8, 7.7 Hz, 1H), 2.29 (dt, J = 14.3, 7.1, 1H), 2.24 (dt, J = 14.3, 7.1 Hz, 1H), 2.11 (dt, J = 14.3, 7.0 Hz, 1H), 1.69 (s, 3H), 1.68 (s, 3H), 1.63 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 145.5, 133.4, 121.5, 112.8, 49.4, 39.9, 25.8, 19.3, 18.0, 11.3.

(+)-2-Benzoyl-3,5-dihydroxy-6-(5-methyl-2-(prop-1-en-2yl)hex-4-en-1-yl)-4,6-bis(3-methylbut-2-en-1-yl)cyclohexa-2,4dienone ((\pm) -24). To a solution of 17 (616 mg, 1.68 mmol) in anhydrous DMF (12 mL) was added NaH (242 mg, 10.1 mmol) at rt. The mixture was stirred at rt for 10 min. Iodide (\pm) -23 (2.66 g, 10.1 mmol) in anhydrous DMF (2 mL) was then added at rt. The reaction mixture was stirred at rt for 1 h. The mixture was quenched with 1 M HCl solution (15 mL) and extracted with EtOAc $(3 \times 20 \text{ mL})$. The combined organics were washed sequentially with H_2O (2 × 30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, $50:1 \rightarrow 15:1$ gradient elution) to give (\pm) -2-benzoyl-3,5-dihydroxy-6-(5-methyl-2-(prop-1-en-2-yl)hex-4-en-1-yl)-4,6-bis(3-methylbut-2-en-1-yl)cyclohexa-2,4-dienone (\pm)-24 (242 mg, 29%, 50% BRSM) as a viscous yellow oil. Data for (\pm)-24: R_f 0.42 (petrol/EtOAc, 4:1); IR (neat) 3278, 2914, 1645, 1445, 1182, 695 cm⁻¹; NMR showed a complex mixture of tautomers and diasteroisomers; HRMS-ESI (m/z) calculated for $C_{33}H_{43}O_4$ [M + H]+ 503.3156, found 503.3149.

 (\pm) -Garcibracteatone (1) and (\pm) -5-epi-Garcibracteatone (25). To a solution of $Mn(OAc)_3(H_2O)_2$ (457 mg, 1.70 mmol) and Cu(OAc)₂(H₂O) (154 mg, 0.85 mmol) in degassed AcOH (2 mL) was added (±)-24 (408 mg, 0.81 mmol) in degassed AcOH (12 mL) at rt. The reaction mixture was stirred at rt for 3 h. The mixture was quenched with H_2O (20 mL) and extracted with EtOAc (3 × 30 mL). The combined organics were washed sequentially with H₂O (50 mL), sat. NaHCO3 solution (50 mL), and brine (50 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO_2 (petrol/EtOAc, 20:1 \rightarrow 15:1 gradient elution) to give (\pm) -garcibracteatone (\pm) -1 (56 mg, 14%) as a white crystalline solid. Data for (\pm) -1: R_f 0.62 (petrol/ EtOAc, 4:1); mp 196-199 °C; IR (neat) cm⁻¹ 3468, 2977, 1736, 1706, 1665, 1601, 1450, 1260, 1111, 765; ¹H NMR (600 MHz, CDCl₃) δ 7.69 (dd, J = 7.6, 1.3 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 5.04 (t, J = 7.2 Hz, 1H), 5.01 (t, J = 7.2 Hz, 1H), 2.83 (s, 1H), 2.66 (dd, J = 9.9, 7.8 Hz, 1H), 2.30-2.23 (m, 1H), 2.27 (d, J = 7.3 Hz, 1H), 2.20 (dd, J = 11.3, 10.1 Hz, 1H), 2.12-2.05 (m, 1H), 2.07 (dd, J = 11.5, 7.8 Hz, 1H), 2.01 (dd, J = 11.8, 7.9 Hz, 1H), 1.89-1.82 (m, 1H), 1.77 (d, J = 13.9)Hz, 1H), 1.68 (d, J = 13.8 Hz, 1H), 1.68 (s, 3H), 1.65 (s, 3H), 1.62 (s, 3H), 1.59-1.55 (m, 1H), 1.57 (s, 3H), 1.55 (s, 1H), 1.46 (s, 3H), 1.35 (s, 3H), 1.10 (s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 213.3, 203.3, 200.3, 150.2, 136.4, 134.3, 133.6, 132.4, 126.9, 126.4, 123.5, 122.9, 118.8, 91.8, 70.2, 69.2, 63.2, 56.9, 56.8, 47.5, 41.5, 37.1, 33.0, 32.5, 29.8, 29.0, 26.2, 25.9, 25.8, 25.1, 18.4, 18.0, 17.9; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ [M + H]⁺ 501.2999, found 501.3002. Further elution gave (\pm) -5-epi-garcibracteatone (\pm) -25 (31 mg, 8%) as a white crystalline solid. Data for 25: R_f 0.58 (petrol/EtOAc, 4:1); mp 212-216 °C; IR (neat) 3487, 2964, 1735, 1706, 1666, 1598, 1455, 1298, 1107, 760 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.70 (dd, J =7.6, 1.2 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.35 (d, J = 7.8 Hz, 1H), 5.11 (t, J = 7.3 Hz, 1H), 5.02 (t, J = 6.9Hz, 1H), 2.81 (s, 1H), 2.62 (dd, J = 9.8, 7.7 Hz, 1H), 2.53-2.47 (m, 1H), 2.33-2.26 (m, 1H), 2.29 (d, J = 8.0 Hz, 1H), 2.20 (dd, J = 11.2, 10.1 Hz, 1H), 2.12 (dd, J = 12.3, 10.9 Hz, 1H), 2.05 (t, J = 9.5 Hz, 1H), 2.03 (d, J = 14.2 Hz, 1H), 1.93-1.87 (m, 1H), 1.72-1.65 (m, 1H), 1.69 (s, 3H), 1.66 (s, 3H), 1.58 (s, 3H), 1.58 (s, 3H), 1.47 (s, 3H), 1.43 (d, J = 14.5 Hz, 1H), 1.38-1.34 (m, 1H), 1.35 (s, 3H), 1.10(s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 213.4, 203.3, 200.2, 150.2, 136.3, 134.6, 133.7, 132.6, 126.9, 126.4, 123.5, 122.8, 119.0, 91.8, 71.0, 69.1, 63.0, 56.2, 52.5, 42.5, 37.2, 35.1, 31.1, 30.2, 29.8, 29.5, 26.2, 25.9, 25.8, 24.8, 22.8, 17.9, 17.8; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ $[M + H]^+$ 501.2999, found 501.3018.

(S)-4-Benzyl-3-((S)-5-methyl-2-(prop-1-en-2-yl)hex-4-enoyl)**oxazolidin-2-one** ((+)-27). To a solution of (-)-26 (9.31 g, 35.9 mmol) in THF (100 mL) was added NaHMDS (1.0 M solution in THF, 39.5 mL, 39.5 mmol) at -78 °C. The mixture was stirred at -78°C for 1 h before addition of prenyl bromide (8.30 mL, 71.8 mmol) dropwise at -78 °C. The resultant mixture was stirred at -78 °C for 6 h. The mixture was quenched with satd NH₄Cl solution (100 mL) and extracted with Et₂O (3 × 100 mL). The combined organics were washed with brine (100 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 10:1 as elutant) to give (S)-4-benzyl-3-((S)-5-methyl-2-(prop-1-en-2-yl)hex-4-enoyl)oxazolidin-2-one (+)-27 (7.78 g, 66%) as a colorless oil. Data for (+)-27: 0.40 (petrol/EtOAc, 4:1); IR (neat) 2919, 1775, 1697, 1206, 701 cm⁻¹; $[\alpha]^{25}_{D}$ (c 1.0, CHCl₃) +102.6°; ¹H NMR (600 MHz, CDCl₃) δ 7.33 (t, J = 7.3 Hz, 2H), 7.29–7.25 (m, 1H), 7.21 (d, J = 7.0Hz, 2H), 5.14 (t, J = 7.2 Hz, 1H), 4.89 (d, J = 7.7 Hz, 2H), 4.69–4.63 (m, 1H), 4.47 (dd, J = 9.0, 5.8 Hz, 1H), 4.17-4.11 (m, 2H), 3.22 (dd, J = 13.4, 3.3 Hz, 1H), 2.78 (dd, J = 13.5, 9.3 Hz, 1H), 2.62 (dt, J = 13.5, 9.3 15.8, 8.3 Hz, 1H), 2.36–2.29 (m, 1H), 1.83 (s, 3H), 1.70 (s, 3H), 1.67 (s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 173.3, 153.0, 143.1, 135.3, 133.8, 129.4, 128.9, 127.3, 121.4, 113.4, 65.7, 55.5, 50.3, 37.8, 29.8, 25.8, 21.3, 17.8; HRMS-ESI (m/z) calculated for $C_{20}H_{26}NO_3$ [M + H]+ 328.1907 found 328.1907.

(S)-5-Methyl-2-(prop-1-en-2-yl)hex-4-en-1-ol ((-)-22). To a solution of (+)-27 (3.26 g, 9.98 mmol) in Et_2O (30 mL) and MeOH (0.53 mL, 12.9 mmol) was added LiBH₄ (283 mg, 12.9 mmol) at 0 °C. The resultant mixture was stirred at rt for 1 h. The mixture was quenched with satd NH₄Cl solution (50 mL) and extracted with Et₂O $(3 \times 50 \text{ mL})$. The combined organics were washed with brine (100) mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/ EtOAc, 6:1 as elutant) to give (S)-3-(iodomethyl)-2,6-dimethylhepta-1,5-diene (-)-22 (975 mg, 63%) as a colorless oil. Data for (-)-22: R_f 0.40 (petrol/EtOAc, 4:1); IR (neat) 3351, 2916, 1646, 1440, 1376, 1038, 888 cm $^{-1}$; [α] 25 _D (c 1.0, CHCl $_{3}$) -2.0; 1 H NMR (600 MHz, CDCl₃) δ 5.08 (tt, J = 7.2, 1.2 Hz, 1H), 4.94–4.91 (m, 1H), 4.82 (d, J= 0.6 Hz, 1H), 3.57 (dt, J = 11.3, 5.8 Hz, 1H), 3.50 (dt, J = 8.7, 3 Hz, 1Hz)1H) 2.28 (qd, J = 7.6, 5.2 Hz, 1H), 2.11 (dt, J = 14.6, 7.3 Hz, 1H), 2.04 (dt, *J* = 14.6, 7.1 Hz, 1H), 1.70 (s, 3H), 1.69 (s, 3H), 1.61 (s, 3H), 1.43 (t, J = 5.3 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 145.4, 132.7, 122.0, 113.1, 63.6, 49.9, 28.4, 25.7, 19.5, 17.8.

(S)-3-(lodomethyl)-2,6-dimethylhepta-1,5-diene ((-)-23). To a solution of PPh₃ (3.68 g, 14.0 mmol) and imidazole (954 mg, 14.0 mmol) in CH₂Cl₂ (70 mL) was added I₂ (3.56 g, 1.40 mmol) at 0 °C. The mixture was stirred at 0 °C for 15 min. (-)-22 (2.06 mL, 13.4 mmol) in CH₂Cl₂ (10 mL) was then added dropwise at 0 °C. The reaction mixture was stirred at rt for 2 h. The mixture was quenched with Na₂S₂O₃ solution (18 g in 100 mL of H₂O) and stirred at rt for 10 min. The layers were separated, and the aqueous layer was extracted with CH2Cl2 (100 mL). The combined organics were washed with brine (100 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (neat petrol as elutant) to give (S)-3-(iodomethyl)-2,6-dimethylhepta-1,5-diene (-)-23 (2.70 g, 77%) as a pale orange oil. Data for (-)-23: R_f 0.70 (neat petrol); IR (neat) 2968, 2913, 1647, 1439, 1375, 1187, 893 cm⁻¹; $[\alpha]^{2\bar{s}}_{D}$ (c 1.0, CHCl₃) -1.8; ¹H NMR (600 MHz, CDCl₃) δ 5.03 (tt, J = 7.9, 1.3 Hz, 1H), 4.91– 4.89 (m, 1H), 4.75 (s, 1H), 3.28 (dd, J = 9.8, 5.8 Hz, 1H), 3.19 (dd, J = 9.8, 7.7 Hz, 1H), 2.29 (dt, J = 14.3, 7.1, 1H), 2.24 (dt, J = 14.3, 7.1)Hz, 1H), 2.11 (dt, J = 14.3, 7.0 Hz, 1H), 1.69 (s, 3H), 1.68 (s, 3H), 1.63 (s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 145.5, 133.4, 121.5, 112.8, 49.4, 39.9, 25.8, 19.3, 18.0, 11.3.

(-)-4-Benzoyl-5-hydroxy-6-((*R*)-5-methyl-2-(prop-1-en-2-yl)-hex-4-en-1-yl)-2,6-bis(3-methylbut-2-en-1-yl)cyclohex-4-ene-1,3-dione ((-)-24). To a solution of 17 (620 mg, 1.70 mmol) in anhydrous DMF (12 mL) was added NaH (164 mg, 10.2 mmol) at rt. The mixture was stirred at rt for 10 min. Iodide (-)-23 (2.70 g, 10.2 mmol) in anhydrous DMF (2 mL) was then added at rt. The reaction mixture was stirred at rt for 1 h. The mixture was quenched with 1 M

HCl solution (15 mL) and extracted with EtOAc (3 × 20 mL). The combined organics were washed sequentially with H₂O (2 × 30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 50:1 \rightarrow 15:1 gradient elution) to give (–)-4-benzoyl-5-hydroxy-6-((R)-5-methyl-2-(prop-1-en-2-yl)hex-4-en-1-yl)-2,6-bis(3-methylbut-2-en-1-yl)cyclohex-4-ene-1,3-dione (–)-24 (252 mg, 29%, 50% BRSM) as a viscous yellow oil. Data for (–)-24: R_f 0.42 (petrol/EtOAc, 4:1); IR (neat) 3278, 2914, 1645, 1445, 1182, 695 cm⁻¹; [a]²⁵_D (c 1.0, CHCl₃) –7.8; NMR showed a complex mixture of tautomers and diasteroisomers; HRMS-ESI (m/z) calculated for C₃₃H₄₃O₄ [M + H]⁺ 503.3156, found 503.3149.

(+)-Garcibracteatone (1) and (-)-5-epi-Garcibracteatone (25). To a solution of $Mn(OAc)_3(H_2O)_2$ (280 mg, 1.04 mmol) and Cu(OAc)₂(H₂O) (95 mg, 0.497 mmol) in degassed AcOH (2 mL) was added 24 (250 mg, 0.497 mmol) in degassed AcOH (8 mL) at rt. The reaction mixture was stirred at rt for 3 h. The mixture was quenched with H_2O (15 mL) and was extracted with EtOAc (3 × 20 mL). The combined organics were washed sequentially with H₂O (30 mL), satd NaHCO₃ solution (30 mL), and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography on SiO_2 (petrol/EtOAc, 20:1 \rightarrow 15:1 gradient elution) to give (+)-garcibracteatone (+)-1 (36 mg, 14%) as a white crystalline solid. Data for 1: R_f 0.62 (petrol/EtOAc, 4:1); mp 196–199 °C; IR (neat) cm⁻¹ 3468, 2977, 1736, 1706, 1665, 1601, 1450, 1260, 1111, 765; $[\alpha]^{25}_{D}$ (c 1.0, CHCl₃) +2.0; ¹H NMR (600 MHz, CDCl₃) δ 7.69 (dd, J = 7.6, 1.3 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 5.04 (t, J = 7.2 Hz, 1H), 5.01 (t, J = 7.2 Hz, 1H), 2.83 (s, 1H), 2.66 (dd, J =9.9, 7.8 Hz, 1H), 2.30-2.23 (m, 1H), 2.27 (d, J = 7.3 Hz, 1H), 2.20(dd, J = 11.3, 10.1 Hz, 1H), 2.12 - 2.05 (m, 1H), 2.07 (dd, J = 11.5,7.8 Hz, 1H), 2.01 (dd, *J* = 11.8, 7.9 Hz, 1H), 1.89–1.82 (m, 1H), 1.77 (d, J = 13.9 Hz, 1H), 1.68 (d, J = 13.8 Hz, 1H), 1.68 (s, 3H), 1.65 (s,3H), 1.62 (s, 3H), 1.59-1.55 (m, 1H), 1.57 (s, 3H), 1.55 (s, 1H), 1.46 (s, 3H), 1.35 (s, 3H), 1.10 (s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 213.3, 203.3, 200.3, 150.2, 136.4, 134.3, 133.6, 132.4, 126.9, 126.4, 123.5, 122.9, 118.8, 91.8, 70.2, 69.2, 63.2, 56.9, 56.8, 47.5, 41.5, 37.1, 33.0, 32.5, 29.8, 29.0, 26.2, 25.9, 25.8, 25.1, 18.4, 18.0, 17.9; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ $[M + H]^+$ 501.2999, found 501.3002. Further elution gave (-)-5-epi-garcibracteatone (-)-25 (21 mg, 8%) as a white crystalline solid. Data for 25: R_f 0.58 (petrol/ EtOAc, 4:1); mp 212-216 °C; IR (neat) 3487, 2964, 1735, 1706, 1666, 1598, 1455, 1298, 1107, 760 cm⁻¹; $[\alpha]^{25}$ _D (c 1.0, CHCl₃) -34.1; ¹H NMR (600 MHz, CDCl₃) δ 7.70 (dd, J = 7.6, 1.2 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.35 (d, J = 7.8 Hz,1H), 5.11 (t, J = 7.3 Hz, 1H), 5.02 (t, J = 6.9 Hz, 1H), 2.81 (s, 1H), $2.62 \text{ (dd, } J = 9.8, 7.7 \text{ Hz, } 1\text{H}), 2.53-2.47 \text{ (m, } 1\text{H}), 2.33-2.26 \text{ (m, } 1\text{H})}$ 1H), 2.29 (d, J = 8.0 Hz, 1H), 2.20 (dd, J = 11.2, 10.1 Hz, 1H), 2.12 (dd, J = 12.3, 10.9 Hz, 1H), 2.05 (t, J = 9.5 Hz, 1H), 2.03 (d, J = 14.2)Hz, 1H), 1.93-1.87 (m, 1H), 1.72-1.65 (m, 1H), 1.69 (s, 3H), 1.66 (s, 3H), 1.58 (s, 3H), 1.58 (s, 3H), 1.47 (s, 3H), 1.43 (d, J = 14.5 Hz,1H), 1.38-1.34 (m, 1H), 1.35 (s, 3H), 1.10 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 213.4, 203.3, 200.2, 150.2, 136.3, 134.6, 133.7, 132.6, 126.9, 126.4, 123.5, 122.8, 119.0, 91.8, 71.0, 69.1, 63.0, 56.2, 52.5, 42.5, 37.2, 35.1, 31.1, 30.2, 29.8, 29.5, 26.2, 25.9, 25.8, 24.8, 22.8, 17.9, 17.8; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ [M + H]⁺ 501.2999, found 501.3018

(\pm)-Ethyl 5-Methyl-2-(prop-1-en-2-yl)hex-5-enoate ((\pm)-28). To a solution of fresh LDA prepared *in situ* from iPr₂NH (4.64 mL, 33.1 mmol) and *n*-BuLi (13.2 mL, 2.5 M in hexanes, 33.1 mmol) in THF (50 mL) was added ethyl 3,3-dimethyl acrylate **20** (2.29 mL, 16.5 mmol) dropwise at -78 °C. The reaction mixture was stirred for 30 min, then 4-iodo-2-methyl-1-butene **12** (4.86 g, 24.8 mmol) was added dropwise, and the mixture stirred for 30 min, then warmed to 0 °C, and stirred for a further 2 h. The reaction was quenched with satd NH₄Cl (50 mL), the product was extracted with EtOAc (3 × 100 mL), and the combined organics washed with H₂O (300 mL) and brine (300 mL), dried over MgSO₄, filtered, and concentrated *in vacuo* to give (\pm)-ethyl 5-methyl-2-(prop-1-en-2-yl)hex-5-enoate (\pm)-28 (2.61 g, 81%) as an orange-yellow oil which was used in the next step

without further purification. Data for (\pm)-28: R_f 0.60 (petrol/EtOAc, 4:1); IR(neat) 2933, 1731, 1648, 1447, 1374, 1151, 1030, 890 cm⁻¹.

¹H NMR (600 MHz, CDCl₃) δ 4.90 (s, 1H), 4.89 (s, 1H), 4.73 (s, 1H), 4.69 (s, 1H), 4.14 (q, J = 7.1 Hz, 2H), 3.11–2.94 (m, 1H), 2.04–1.91 (m, 2H), 1.75 (s, 3H), 1.72 (s, 3H), 1.76–1.67 (m, 2H), 1.25 (t, J = 7.1 Hz, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 173.6, 145.0, 142.4, 113.8, 110.5, 60.5, 52.5, 35.4, 28.0, 22.3, 20.2, 14.2; HRMS-ESI (m/z) calculated for C₁₂H₂₁O₂ [M + H]⁺ 197.1536, found 197.1534.

 (\pm) -5-Methyl-2-(prop-1-en-2-yl)hex-5-en-1-ol $((\pm)$ -29). To a solution of LiAlH₄ (1.11 g, 29.3 mmol) in Et₂O (35 mL) at 0 °C was added a solution of (\pm) -28 (2.61 g, 13.3 mmol) in Et₂O (20 mL) dropwise over 10 min. The reaction mixture was warmed to rt and stirred for 1 h and then quenched carefully with successive addition of H₂O (1.11 mL), 15% NaOH (1.11 mL), and H₂O (3.33 mL). The product was extracted with EtOAc (2 × 30 mL), the undesired precipitate was removed from the combined organics by vacuum filtration, the precipitate was washed thoroughly with EtOAc (4 × 30 mL), and the filtrate was concentrated in vacuo. The resultant residue was purified by flash chromatography (SiO2, petrol/EtOAc, 10:1) to give (\pm) -5-methyl-2-(prop-1-en-2-yl)hex-5-en-1-ol (\pm) -29 (1.89 g, 92%) as a clear oil. Partial data for (\pm) -29: R_f 0.30 (petrol/EtOAc, 4:1); IR (neat) 3347, 3074, 2934, 1647, 1447, 1374, 1042, 885 cm⁻¹. ¹H NMR (600 MHz, CDCl₃) δ 4.96 (s, 1H), 4.84 (s, 1H), 4.71 (s, 1H), 4.67 (s, 1H), 3.60-3.41 (m, 2H), 2.33-2.23 (m, 1H), 2.03-1.91 (m, 2H), 1.72 (s, 3H), 1.69 (s, 3H), 1.54-1.42 (m, 2H), 1.39 (t, J = 1.42 (m, 2H), 1.39 (t, J = 1.42 (m, 2H), 1.42 (m, 2H), 1.44 (m, 2H), 1.5.6, 1H); 13 C NMR (150 MHz, CDCl₃) δ 145.6, 144.8, 114.1, 110.0, 64.0, 49.5, 35.3, 27.2, 22.5, 18.7; HRMS-ESI (m/z) calculated for C₁₀H₁₉O [M + H]⁺ 155.1430, found 155.1429.

(+)-3-(lodomethyl)-2,6-dimethylhepta-1,6-diene ((+)-30). To a solution of PPh₃ (5.70 g, 21.8 mmol) and imidazole (1.48 g, 21.8 mmol) in CH₂Cl₂ (100 mL) at 0 °C was added I₂ (5.52 g, 21.8 mmol) portion wise and stirred for 15 min. (\pm)-29 (3.05 g, 19.8 mmol) in CH₂Cl₂ (20 mL) was added dropwise, then the reaction mixture was warmed to rt and stirred for 2.5 h. The reaction was quenched by with Na₂S₂O₃ solution (11 g in 60 mL of H₂O) and stirred for 10 min, then the product was extracted with CH₂Cl₂ (3 × 50 mL), and the combined organics were washed with brine, dried over MgSO₄, filtered, and concentrated in vacuo. The resultant residue was purified by flash chromatography (SiO2, neat petrol) to give (\pm) -3-(iodomethyl)-2,6-dimethylhepta-1,6-diene (\pm) -30 (3.71 g, 71%) as a pale orange oil. Data for (\pm) -30: R_f 0.85 (petrol/EtOAc, 4:1); IR (neat) 3074, 2935, 1647, 1445, 1374, 1183, 888 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 4.92 (s, 1H), 4.78 (s, 1H), 4.72 (s, 1H), 4.68 (s, 1H), 3.21 (ddd, J = 17.6, 9.8, 7.2 Hz, 2H), 2.32 (m, 1H), 2.02-1.91 (m, 2H), 1.72 (s, 3H), 1.74-1.68 (m, 1H), 1.66 (s, 3H), 1.55–1.48 (m, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 145.1, 144.9, 113.6, 110.2, 48.9, 35.4, 30.8, 22.4, 18.4, 11.4.

 (\pm) -2-Benzoyl-3,5-dihydroxy-6-(5-methyl-2-(prop-1-en-2yl)hex-4-en-1-yl)-4,6-bis(3-methylbut-2-en-1-yl)cyclohexa-2,4dienone ((±)-31). NaH (566 mg, 23.6 mmol) was added portionwise to a solution of 17 (1.24 g, 3.38 mmol) in DMF (30 mL) at rt and stirred for 10 min. Iodide (\pm) -30 (5.63 g, 21.3 mmol) was added dropwise, and the reaction mixture was stirred for 30 min before being quenched with satd NH₄Cl (30 mL). The product was extracted with EtOAc (4 × 50 mL), and the combined organics were washed with brine (5 × 100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. The resultant residue was purified by flash chromatography (SiO₂, petrol/EtOAc, 15:1 \rightarrow 5:1) to give (±)-2-benzoyl-3,5dihydroxy-6-(5-methyl-2-(prop-1-en-2-yl)hex-4-en-1-yl)-4,6-bis(3methylbut-2-en-1-yl)cyclohexa-2,4-dienone (\pm)-31 (554 mg, 33%) as a viscous yellow oil. Partial data for (\pm) -31: R_f 0.20 (petrol/EtOAc, 4:1); IR(neat) 3408, 2967, 2929, 1662, 1560, 1447, 1376, 909, 888, 731, 696 cm⁻¹ NMR spectra showed a complex mixture of tautomers and diastereoisomers; HRMS-ESI (m/z) calculated for C₃₃H₄₃O₄ [M + H]+ 503.3156, found 503.3156.

(\pm)-Doitunggarcinone A (2) and (\pm)-5-epi-Doitunggarcinone A (32). To a solution of Mn(OAc)₃(H₂O)₂ (591 mg, 2.20 mmol) and Cu(OAc)₂(H₂O) (210 mg, 1.10 mmol) in degassed AcOH (10 mL) was added dropwise a solution of (\pm)-31 (554 mg, 1.10 mmol) in degassed AcOH (7 mL) at rt. The reaction mixture was

stirred for 3 h and quenched with H₂O (15 mL), and the product was extracted with EtOAc (4 × 50 mL). The combined organics were washed with H_2O (50 mL), satd NaHCO₃ (2 × 100 mL), and brine (100 mL), then dried over MgSO₄, filtered, and concentrated in vacuo. The resultant residue was purified by flash chromatography (SiO₂, petrol/EtOAc, 20:1) to give (\pm) -doitunggarcinone A (\pm) -2 (71 mg, 13%) as an off-white powder. Data for (\pm) -2: R_f 0.53 (petrol/EtOAc, 4:1); mp 221.8-226.4 °C; IR (neat) 3495, 2935, 1736, 1707, 1668, 1450, 1313, 1300, 1255, 1109, 885, 767 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.69 (dd, J = 7.6, 1.4 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (td, I = 7.6, 0.8 Hz, 1H), 7.35 (d, I = 7.6 Hz, 1H), 5.04 (t, I = 7.2Hz, 1H), 4.70 (s, 1H), 4.67 (s, 1H), 2.87 (s, 1H), 2.67 (dd, J = 9.9, 7.8Hz, 1H), 2.27 (d, J = 7.1 Hz, 2H), 2.21 (dd, J = 13.4, 11.5 Hz, 1H), $2.08 \text{ (dd, } I = 13.4, 11.5 \text{ Hz, } 1\text{H}), 2.09-2.01 \text{ (m, } 2\text{H}), 1.86-1.77 \text{ (m, } 2\text{H})}$ 2H), 1.77 (d, J = 14.0 Hz, 1H), 1.75-1.68 (m, 1H), 1.71 (s, 3H), 1.66(d, J = 14.0 Hz, 1H), 1.65 (s, 3H), 1.61-1.55 (m, 1H), 1.57 (s, 3H),1.49 (m, 1H), 1.44 (s, 3H), 1.36 (s, 3H), 1.09 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 213.2, 203.2, 200.2, 150.1, 145.6, 136.4, 134.4, 133.7, 127.0, 126.4, 123.5, 118.8, 110.1, 91.8, 70.2, 69.2, 63.2, 56.8, 56.4, 47.5, 41.6, 37.2, 36.5, 32.8, 32.6, 29.7, 29.0, 26.2, 25.9, 25.1, 22.5, 18.4, 17.9; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ [M + H]⁺ 501.2999, found 501.3001. Further elution gave (\pm) -5-epi-doitunggarcinone A (\pm) -32 (69 mg, 12%) as an off-white powder. Data for (\pm) -32: R_f 0.50 (petrol/EtOAc, 4:1); mp 210.0-214.0 °C; IR (neat) 3480, 2924, 1734, 1704, 1669, 1452, 1314, 1103, 898, 763 cm⁻¹. ¹H NMR (600 MHz, CDCl₃) δ 7.70 (dd, J = 7.6, 1.3 Hz, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.37 (dt, J = 7.5, 1.8 Hz, 1H), 7.35 (d, J = 7.8 Hz, 1H), 5.08(t, J = 7.3 Hz, 1H), 4.71 (s, 1H), 4.65 (s, 1H), 2.82 (s, 1H), 2.62 (dd, J= 9.9, 7.7 Hz, 1H), 2.45 (tt, J = 11.8, 3.5 Hz, 1H), 2.28 (d, J = 8.0 Hz,2H), 2.22 (dd, *J* = 11.3, 10.0 Hz, 1H), 2.16 (dd, *J* = 12.3, 10.7 Hz, 1H), 2.07 (dd, J = 11.4, 7.7 Hz, 2H), 2.01 (d, J = 14.4 Hz, 1H), 1.95-1.89(m, 1H), 1.69 (s, 3H), 1.65 (s, 3H), 1.58 (s, 3H), 1.45 (s, 3H), 1.44 (d, J = 14.4 Hz, 1H), 1.45–1.33 (m, 2H), 1.36 (s, 3H), 1.10 (s, 3H) 1.08–1.02 (m, 1H); 13 C NMR (150 MHz, CDCl₃) δ 213.4, 203.2, 200.2, 150.2, 145.2, 136.4, 134.6, 133.7, 127.0, 126.4, 123.5, 118.9, 110.2. 91.7, 71.1, 69.0, 62.9, 56.2, 51.7, 42.5, 37.2, 36.8, 35.1, 31.3, 29.8, 29.7, 29.5, 26.2, 25.8, 24.8, 22.6, 22.4, 17.9; HRMS-ESI (m/z) calculated for $C_{33}H_{41}O_4$ [M + H]⁺ 501.2999, found 501.3001.

Phenyl(2,4,6-trihydroxy-3,5-diisopentylphenyl)methanone (33). To a solution of 17 (1.20 g, 3.27 mmol) in EtOH (40 mL) was added Pd/C (50 mg) at rt. The flask was evacuated three times and placed under an atmosphere of H2. The reaction mixture was stirred at rt for 2 h. The mixture was filtered through a pad of Celite extracting with EtOAc, and the filtrate was concentrated in vacuo. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 6:1 as elutant) to give phenyl(2,4,6-trihydroxy-3,5-diisopentylphenyl)methanone 33 (950 mg, 79%) as a yellow crystalline solid. Data for 33: R_f 0.49 (petrol/EtOAc, 4:1); IR (neat) 3505, 2949, 1621, 1561, 1314, 1191, 1092, 926, 698 cm⁻¹; mp 82–83 °C; ¹H NMR (600 MHz, CDCl₃) δ 8.80 (s, 1H), 7.64 (d, J = 7.1 Hz, 2H), 7.59 (t, J = 7.4 Hz, 1H), 7.53 (t, J = 7.6 Hz, 2H), 5.40 (s, 1H), 2.54 (dt, J = 14.7, 6.0 Hz, 4H), 1.65-1.55 (m, 2H), 1.40-1.34 (m, 4H), 0.95 (s, 3H), 0.94 (s, 3H); 13 C NMR (150 MHz, CDCl₃) δ 197.6, 159.6, 157.4, 139.9, 132.3, 129.4, 127.8, 107.8, 104.4, 38.1, 28.2, 22.5, 20.7; HRMS-ESI (m/z) calculated for $C_{23}H_{31}O_4$ $[M + H]^+$ 371.2217, found 371.2221.

2-Benzoyl-3,5-dihydroxy-4,6-diisopentyl-6-(3-methylbut-3-en-1-yl)cyclohexa-2,4-dienone (34). To a solution of 33 (409 mg, 1.10 mmol) in anhydrous DMF (10 mL) was added NaH (80 mg, 3.3 mmol) at rt. The mixture was stirred at rt for 5 min before 4-iodo-2-methyl-1-butene **12** (0.26 mL, 2.20 mmol) was then added at rt. The reaction mixture was stirred at rt for 1 h. The mixture was quenched with 1 M HCl solution (10 mL) and extracted with EtOAc (3 × 15 mL). The combined organics were washed sequentially with $\rm H_2O$ (2 × 30 mL) and brine (30 mL), dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on SiO₂ (petrol/EtOAc, 8:1) to give 2-benzoyl-3,5-dihydroxy-4,6-diisopentyl-6-(3-methylbut-3-en-1-yl)cyclohexa-2,4-dienone 34 (177 mg, 37%) as an off white solid. Data for 34: R_f 0.28 (petrol/EtOAc, 4:1); mp 116–119 °C; IR (neat) 3187, 2956, 1646, 1447, 1207, 1171, 692 cm⁻¹; NMR spectra showed a complex mixture of

tautomers; HRMS-ESI (m/z) calculated for $C_{28}H_{39}O_4$ $[M + H]^+$ 439.2843, found 439.2847.

ASSOCIATED CONTENT

S Supporting Information

 1 H and 13 C NMR spectra for compounds 1, 2, 10, 11, 13, 16–19, 21–25, and 27–34 and chiral HPLC traces for (+)-1, (-)-1, and (\pm)-1. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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